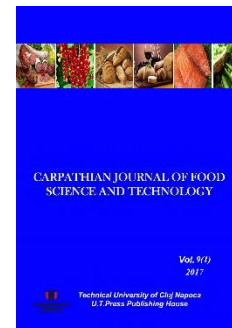


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ANTIBACTERIAL ACTIVITY OF VARIOUS EXTRACTS FROM *THYMUS TRANSCAPICUS* AGAINST FOOD PATHOGENIC MICROORGANISMS

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ABSTRACT

Keywords:

Thymus transcasicus;
Antimicrobial activity;
Extract;
MIC and MBC;

Thyme (*Thymus transcasicus*) is a herbaceous perennial plant belonging to the *Lamiaceae* family. Thymus species are considered as medicinal plants due to their pharmacological and biological properties. In the present study, we have investigated the antibacterial capabilities of dichloromethane, ethylacetate and methanol extracts of *Thymus transcasicus*. Antibacterial activities was screening against two Gram-positive bacteria (*Staphylococcus aureus*, and *Listeria Monocytogenes*) and two Gram-negative bacteria (*Salmonella enterica*, *Escherichia coli*) by detection of minimum inhibitory concentration (MIC), minimum bactericidal concentration (MBC) method. The highest antibacterial index was observed from methanol extract on *Staphylococcus aureus* in MIC-MBC method ($p < 0.05$). The results of this investigation indicated that *Thymus transcasicus* was found to possess moderate antibacterial activities. Further research is required to identify the active photochemical responsible for these biological activities.

1. Introduction

Treatment of infectious diseases continues to be problematic in modern time. With the available antiviral and antimicrobial drugs, the treatment often leads to the problem of resistance (Vijayan et al. 2004; Mehrgana et al. 2008). There is little likelihood that available orthodox antiviral drugs can eliminate all or even most viral diseases (Jassim & Naji 2003). The search for new antimicrobial substances exhibiting minimal side effects is warranted because of the severe side effects of some drugs currently in use (Mehrgana et al. 2008; Kandil et al. 1994). So, there is an increasing need for new substances with antiviral and antimicrobial activity. Medicinal plants have been traditionally used for different kinds of ailments including infectious diseases (Vijayan et al. 2004). Therefore, the development of new medicinal plant products is vital in controlling

the threats posed by some pathogenic microorganisms (Jassim & Naji 2003).

Lamiaceae (formerly Labiatae) is one of the most important plant families in which *Thymus* with about 215 species, is a significant genus (Mehrgana et al. 2008). *Thymus* species are well known as medicinal plants because of their biological and pharmacological properties. In traditional medicine, the extracts of different species of *Thymus* have been widely used for the treatment of gastritis, diarrhea and enuresis in children, bronchitis and whooping cough (Pertussis) (Ebadi 2002).

Several studies have been shown that *Thymus* species have antibacterial (Mehrgana et al. 2008; Figueiredo et al. 2008; Tohidpour et al. 2010), antifungal (Figueiredo et al. 2008. Bonjar 2004; Sokovic et al. 2009), cytotoxic (Goncalvesa et al. 2010), analgesic (Sokovic et al. 2009), antiparasitic (Goncalvesa et al. 2010), topical anti-inflammatory (Ismaili et al.

43 2002), antispasmodic (Begrow et al. 2009),
 44 mosquitocidal (Pavela et al. 2009) and
 45 antioxidant (Zamani et al. 2009; Soares et al.
 46 1997) activities. Antiviral effect of extracts
 47 from some plants of the Lamiaceae family
 48 against HSV-1 (Vijayan et al. 2004; Nolkemper
 49 et al. 2006) and HSV-2 (Nolkemper et al. 2006)
 50 has been reported and the extract of *Thymus*
 51 *vulgaris* has been shown antiviral activity
 52 (Nolkemper et al. 2006).

53 Antioxidant (Zamani et al. 2009) and
 54 antiemetic (Moallem et al. 2009) effects of
 55 *Thymus transcaspicus* have been reported in
 56 some studies.

57 The aim of this study was to evaluate the
 58 antimicrobial activities of the dichloromethane,
 59 ethylacetate and methanol extracts of *Thymus*
 60 *transcapicus*. To the best of our knowledge, we
 61 are the first to report that *Thymus transcaspicus*
 62 extracts showed antimicrobial activity against
 63 the pathogenic microorganisms that were
 64 tested.

66 2. Materials and methods

68 2.1. Plant material

69 The Plant material was collected in May
 70 2016 from North Khorasan Province Mountains
 71 in Iran. Then, the plant was identified and
 72 confirmed by Natural Products & Medicinal
 73 Plants Research Centre, North Khorasan
 74 University of Medical Sciences (Iran) and
 75 Voucher specimen (No: MP 32/4) was
 76 deposited in herbarium of the Natural Products
 77 & Medicinal Plants Research Centre.

79 2.2. Preparation of plant extracts

81 The aerial parts of the plants were dried
 82 under shade at room temperature and then cut
 83 into small pieces. About 100 g of sample was
 84 macerated in methanol, dichloromethane, and
 85 ethylacetate at room temperature for 48 h
 86 separately. Each solvent was allowed to remain
 87 in contact with plant material for 24 h, and
 88 replaced with fresh solvent four times.
 89 Removal of the solvents under vacuum at 40 °C
 90 gave the crude extracts (Boozari et al. 2015).

92 2.3. Antimicrobial Activity

93
 94 Determination of the minimum inhibitory
 95 concentrations (MIC) antimicrobial activities of
 96 methanol, dichloromethane, and ethylacetate
 97 extracts of the aerial part of the plant were
 98 determined against two Gram-positive bacteria:
 99 *S. aureus* (ATCC 6538p), and *L.*
 100 *monocytogenes* (ATCC 35152), two Gram-
 101 negative bacteria: *S. enterica* (ATCC 53648),
 102 *Escherichia coli* (ATCC 10536).

104 2.5. Determination of the minimal inhibitory 105 concentration (MIC)

106
 107 Minimum inhibitory concentrations (MIC)
 108 were determined by broth macro dilution
 109 method in 96-well plates by Rios and Duffy
 110 methods (Rios et al. 1988; Duffy & Power
 111 2001).

112 Initial concentration of each extract was
 113 prepared with the aid of bath sonicator (0.8 g
 114 extract with 4 ml solvent and 30% dimethyl
 115 sulphoxide in sterile distilled water and one
 116 drop of Tween 80). 1 ml of diluted extract was
 117 infused into macro-plate with 1ml of sterile
 118 Mueller-Hinton broth (MHB; Hi Media, India)
 119 and then diluted (50% with MHB). 0.5
 120 McFarland standard turbidity for microbial
 121 suspension equivalent was prepared by
 122 suspensions of the growth from brain-heart
 123 infusion medium (HiMedia, India).
 124 Suspensions were further diluted to obtain a
 125 concentration of 10⁷ colony-forming units
 126 (CFU) per ml for the bacteria. Then, 10 µl of
 127 diluted inoculums was added to each well of
 128 macro-plate. The sterility of the medium was
 129 also tested in two wells and Gentamicin was
 130 used as the positive control for bacterial strains.
 131 Plates were incubated for 24 h at 37°C for
 132 bacteria. The growth of microorganisms was
 133 assessed by TTC (2, 3, 5-triphenyl tetrazolium
 134 chloride, Sigma, USA) assay. Briefly, 0.5 ml of
 135 TTC (5 mg/ml; dissolved in sterile water) was
 136 added to each well and the plates were
 137 incubated at 37 °C for bacteria. The results
 138 were expressed as the lowest concentration of
 139 plant extract that could inhibit any red dye
 140 production. MIC values were defined as the
 141 lowest concentrations of oil that inhibit bacteria

142 after 24 h. All experiments were done in
143 triplicates.

144

145 **2.6. Determination of minimum bactericidal** 146 **concentrations (MBC)**

147

148 The bactericidal effects of extracts were
149 determined according to the method described
150 by Rios (Rios et al. 1988). 100 µl of clear
151 dilutions in wells of macro-plate were sub
152 cultured on the Mueller- Hinton agar plates and
153 subsequently incubated at 37 °C for 24 h.
154 Minimal bactericidal concentration (MBC)
155 were recorded from the first tube that showed
156 no growth on solid media. All experiments
157 were done in triplicates.

158

159 **2.7. Statistical analysis**

160

161 Data for each test were statistically
162 analyzed. The statistical analysis of the data
163 was performed using SPSS statistical software
164 version 18 (SPSS Inc., Chicago, IL). Data
165 recorded as means ± standard deviation of three
166 replicate measurements.

167

168 **3. Results and Discussion**

169

170 The highest yield of extract was for
171 methanol extract with 2.92 %, dichloromethane
172 and ethylacetate extracts had 1.78 % and 1.58
173 % yields.

174 To our knowledge, this is the first report on
175 the antimicrobial activity of *Thymus*
176 *transcapicus*. Experimental studies carried out
177 in species of *Thymus* have identified phenols
178 and flavonoids as phytochemicals with
179 antimicrobial properties. The *Thymus*
180 *transcapicus* extracts were tested for
181 antimicrobial activity against 4 different
182 pathogenic microorganisms, including 2 Gram-
183 positive and 2 Gram-negative bacteria.

184 The Gram-positive strains of bacteria that
185 were tested seemed to be more sensitive to the
186 extracts, which are attributed to the absence of
187 an outer lipopolysaccharide layer in Gram-
188 negative bacteria that provides a resistant
189 barrier (Inouye et al. 2001).

190 The antibacterial activity of flavonoids and
191 polyphenols has been attributed to inhibition of

192 synthesis of RNA and DNA (Arora et al. 2013).
193 Thus, the antibacterial activity of ethylacetate
194 and methanol extracts of *Thymus transcapicus*
195 could be attributed to the high polyphenolic
196 compounds present in the extract.

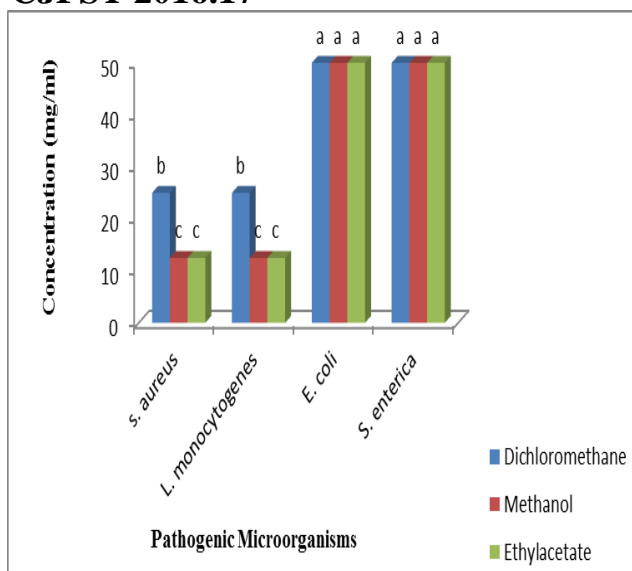
197 The negative control (DMSO) is showed
198 activity against all the microbial strains tested.
199 The positive control was showed activity
200 against all the microbial strains. The extracts
201 can inhibit the growth of microbial strains the
202 growth inhibitory effects of the *Thymus*
203 *transcapicus* extract were concentration
204 dependent (He et al. 2010).

205 The standard antibiotic was more potent,
206 having lower MIC values against bacteria. The
207 minimum inhibitory concentrations (MIC) of
208 different extracts were determined by preparing
209 solution of varying concentration (12.5- 100
210 mg/ml). The methanol and ethylacetate extracts
211 exhibited antibacterial properties against Gram-
212 positive tested bacteria.

213 According to the present results, these
214 extracts did not show any inhibitory activity
215 against all the Gram-negative microorganisms
216 and Gram-positive bacteria were more sensitive
217 to these extracts than Gram-negative bacteria.

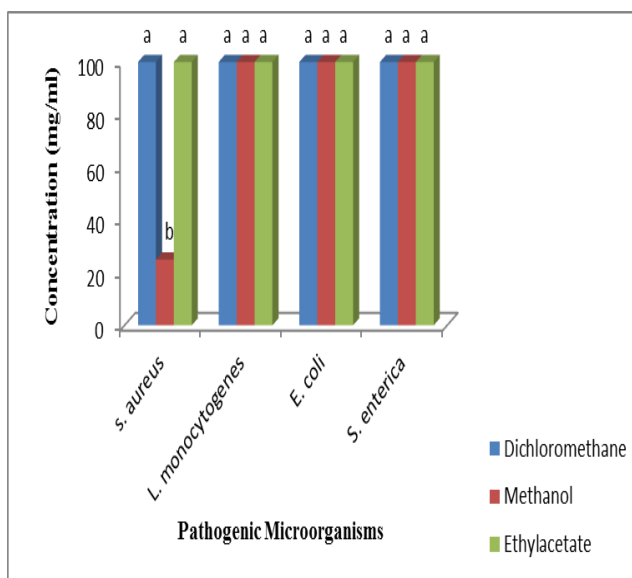
218 As shown in Figs 1 and 2, the results
219 indicated that among the three extracts,
220 ethylacetate extract has greater antimicrobial
221 activity against the tested microorganisms
222 compared to dichloromethane and methanol
223 extract.

224 Methanol extract showed good activity
225 against *S. aureus*, showing the inhibition the
226 lowest MIC values (12.5 mg/ml) and the lowest
227 MBC values (25 mg/ml) ($p < 0.05$).



228
229

230 **Figure 1.** Comparison of minimum inhibitory
231 concentration (MIC, mg/ml) of methanol,
232 dichloromethane and ethyl acetate extracts of
233 *Thymus transcaspicus*.
234 (Each experiment was tested in triplicate.)
235



236
237

238 **Figure 2.** Comparison of minimum bactericidal
239 concentrations (MBC, mg/ml) of methanol,
240 dichloromethane and ethyl acetate extracts of
241 *Thymus transcaspicus*
242 (Each experiment was tested in triplicate.)
243

244 Antimicrobial activity of the plants of
245 different areas of the world has been reported
246 (Janovska et al, 2003).

247 Recent studies have shown that *Thymus*
248 species have antibacterial, antifungal, and

249 antioxidant activities (Rahimmalek *et al.*, 2009;
250 Jordan *et al.*, 2009; Bassam *et al.*, 2004). The
251 anti-bacterial characteristic of *Thymus* spp. is
252 due to the occurrence of thymol in this genus.
253 This substance can be used as a disinfectant.
254 Previous studies showed that the essential oil
255 and extract from *T. daenensis* exhibited
256 antimicrobial activities against *Candida*
257 *albicans* (Ghasemi Pirbalouti *et al.*, 2009a),
258 *Listeria monocytogenes* (Ghasemi Pirbalouti *et*
259 *al.*, 2009b), *Campylobacter jejuni* and
260 *Campylobacter coli* (Ghasemi Pirbalouti *et al.*,
261 2010a), *Staphylococcus aureus*, *Escherichia*
262 *coli*, *Pseudomonas aeruginosa*, *Klebsiella*
263 *pneumoniae* (Ghasemi Pirbalouti *et al.*, 2010b),
264 *Escherichia coli* O157:H7 (Ghasemi Pirbalouti
265 *et al.*, 2010c), and *Saprolegnia parasitica*
266 (Ghasemi Pirbalouti *et al.*, 2009c).

267 Antimicrobial activities of some *Thymus*
268 species have been shown in other previous
269 studies. *Thymus pubescens* and *Thymus*
270 *vulgaris* extract demonstrated good
271 antibacterial activity against some drug
272 resistant Gram-positive bacteria (Mehrgana *et*
273 *al.*, 2008; Tohidpour *et al.*, 2010). The essential
274 oil of the *Thymus caramanicus* showed high
275 inhibitory activity against *Helicobacter pylori*
276 (Eftekhar *et al.*, 2009). *T. transcaspicus*
277 essential oil was tested for its antibacterial
278 activity against various Gram-positive and
279 Gram-negative bacteria Standard strains. All
280 the bacteria were inhibited by the essential oil
281 but in variable degrees.

282 Inhibition of *Staphylococcus aureus* (de
283 Oliveira *et al.*, 2010) and antibacterial effects
284 against *E. coli* (Pei *et al.*, 2009) by thymol and
285 carvacrol have been reported. Carvacrol also
286 has been reported to exhibit a dose dependent
287 inhibitory effect on *Vibrio cholerae* in food
288 (Rattanachaikunsopon *et al.*, 2010). Thymol,
289 which is the main component of many *Thymus*
290 spp. and also in the oil of *T. transcaspicus*
291 (64%), is known as an antiseptic agent (Miri *et*
292 *al.*, 2002).

293 Several studies have demonstrated the
294 antimicrobial activity the essential oils and/or
295 the extracts of many species of the genus
296 *Thymus* rich in volatile phenols and/or volatile
297 alcohols (Sokovic *et al.* 2009; Laouer *et al.*

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