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NUTRACEUTICALS COMPOUNDS EXTRACTION OPTIMIZATION FROM OPEN AIR AND SWELL-DRIED BANANA PEEL POWDERS

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Article history:	ABSTRACT
Received	The aim of this study was to optimize the operating conditions of two
13 September 2022	drying processes on banana peels: open air and Instant Controlled Pressure
Accepted	Drop technique (DIC) assisted Swell-Drying at 40°C in order to obtain
29 November 2022	high quality final powders. The optimization of three extraction conditions
Published	including extraction temperature (40-100°C), extraction time $(10 - 60 \text{ min})$
December 2022	and particle size $(60 - 363 \ \mu m)$ from open air banana peel powder was
Keywords:	investigated). Additionally, three DIC texturing conditions were improved.
Banana peel;	DIC involves maintaining banana peels at a high temperature for 20 to
Open air drying;	220s, high steam pressure (p=0.3 to 0.6 MPa) and varying the Number of
DIC Swell-Drying;	cycles from 1 to 7. Modelling of some nutraceutical compounds (Total
Optimization;	Flavonoids Content and carotenoids) from open air and Swell-Dried
Nutraceutical compounds;	banana peels powders by applying experiments design using Response
Modelling	Surface Methodology and Desirability Function. The antioxidant activity
	was also investigated by the determination of the % of DPPH inhibition.
	The optimal conditions derived from the multi-Responses-Desirability
	Function were as follows: 60.47°C; 10min; and particle size Φ = 348.648
	μ m yielding a TFC=5.13 (mg QE/g d.b), TCC=0.48 (mg /g d.b) and % of
	DPPH inhibition=73.05%, with an optimal desirability coefficient d=0.7
	(open air process). The following optimized DIC operating parameters
	with maximum desirability coefficient d=1, t=24.46s P=0.59MPa and
	number of cycles N=6.38 yielding a TFC=4.07 (mg QE/g d.b), a TCC=
	1.37 (mg /g d.b) and a % of DPPH inhibition=75.97 %. Banana peel could
	be a good source of bioactive substances, which could be further used as a
	natural antioxidant.

1.Introduction

The production of banana fruit, a tropical perishable fruit, amounts to 119 million tonnes per year (FAO, 2022), with around 36 million tonnes of peel as a waste product. Banana peel

contains a variety of bioactive compounds, including flavonoids, tannins, carotenoids, vitamins, and other elements with antiinflammatory and antioxidant properties (Havsteen, 1983; Toh et *al.*, 2016; Pereira et *al.*, 2017). According to (Iman and Akter, 2011), the eucocyanidin flavonoid is gaining popularity as medication for stomach ulcers.

Banana peel contains mucilages, which are naturally occurring polysaccharides that are chemically linked to proteins and minerals (Gemede et *al.*, 2015). These substances have a range of medicinal effects including anticancer, antioxidant, antimicrobial, hypoglycemic, and antiulcer properties (Dantas et *al.*, 2021).

Proteins and other compounds have the ability to form phenolic insoluble complexes. Dopamine is the only phenolic amino acid found in banana peels according to Happy (Emaga et *al.*, 2007).

In terms of applications, the banana's peel has long been used in traditional medicine to cure many health disorders such as burns, anemia, and diarrhoea (Vu et *al.*, 2018).

In Algerian traditional medicine, dried banana peel infusion is used to cure gastric disorders.

Furthermore, a water extract of *Musa* acuminata peel dried in the open air could be deemed an efficient treatment for reducing gastrointestinal inflammation in rats after 30 days. This work was conducted at Pasteur Institute in Algeria and is currently being published.

The impact of pre-treatment on the availability of phenolic chemicals recovered

from banana peels have been reported in a number of studies. These pre-treatments include, among others, blanching (Hernandez-Carranza, 2016), and freezing (ISO, 2000).

To our knowledge, no scientific research has been conducted to investigate the impact of the two drying processes -open air drying and Swell-Drying using DIC- on the availability of nutraceutical compounds of banana peel and determine the parameters under which it can be used.

The current research was carried out in order to analyse the biochemical composition of banana peels and to optimize the extraction conditions of some nutraceutical compounds (flavonoids and carotenoids) and to evaluate their antioxidant activity by determination of the % of DPPH inhibition from open air and DIC Swell-dried banana peels powders by applying experiments design using Response Surface Methodology (RSM) and Desirability Function (DF).

2. Materials and methods

2.1. Plant material

Banana fruits of the *Musa acuminate* species were acquired at a French market in La Rochelle (France). Table 1 summarizes some physicochemical properties of fresh Musa acuminate peels.

Parameters	Average contents						
pH at 22°C	5.45±0.07						
Humidity level (%)	7.76±0.18						
Ash rate (%)	14.18±4.14						
Organic matter (%)	85.82±4.14						
Titratable acidity (g of citric acid /100g of sample)	0.09±0.01						
Total sugars (g/l)	17.11±0.02						
Reducing sugars (g/l)	0.36±0.01						
Sucrose (g/l)	15.91±1.64						

 Table 1. Some physicochemical composition of fresh Musa acuminate peels (n=3)

2.2. Methods

2.2.1. Drying processes of banana peels

Two drying processes were used in this study: open air under dark condition and Instant Controlled Pressure Drop process DIC assisted Swell-Drying at 40°C. This work was conducted at the Unit Laboratory of Materials, Processes & Environment (UR-MPE), M'Hamed Bougara University of Boumerdes (Algeria) with the collaboration of the laboratory of Engineering Science for Environment (LaSIE) UMRER7356 CNRS, La Rochelle University (France).

All banana fruits were washed under tap water to remove impurities, then the peels were manually recovered and cut into small pieces before being dried in the open air under dark condition and textured with DIC using equipment from ABCAR-DIC Process (La Rochelle, France) followed by Swell-Drying at 40°C until the final weight was constant.

The DIC treatment involves four steps, which are as follows: 1): a primary vacuum is created; 2): high saturated steam pressure is injected and maintained for a short time (10 to 30 s); 3): pressure is abruptly dropped toward a vacuum within few milliseconds, and finally released to atmospheric pressure.

The dried peels were crushed in an electric grinder and sieved using an ORTO ALRESA sieve between 60 and 363 μ m. The final powders were stored in hermetic glass bottle at room temperature until their analysis

2.2.2. Experimental design

Using a banana peel that had been dried in the open air and DIC assisted Swell-Drying, the extraction of nutraceutical compounds (Total of flavonoids and carotenoids) and antioxidant activity were improved as revealed by the response optimizer function of Statgraphics Centurion 18 software. The independent variables used in the RSM design of a banana peel that had been dried in the open air are listed in Table 2. Three DIC operating conditions optimization of banana peel: treatment time (X_1) from 20 to 220s, steam pressure P (X_2) from 0.3 to 0.6 MPa, and the number of cycles (X_3) adjusted from 1 to 7 were also investigated (Table 3).

The response for nutraceutical components (TFC and TCC) and the percentage of DPPH inhibition of water extract from banana peels dried in open air and DIC-assisted Swell-Drying were assessed using a Central Composite Design (CCD) with four central points (Tables 4, 5).

A second-order polynomial of the following form was employed to express the responses using multiple regressions:

$$Y = a_0 + \sum_{i=1}^{3} a_i X_i + \sum_{i=1}^{3} a_i X_i^2 + \sum_{i=1}^{3} \sum_{j=1}^{3} a_{ij} X_i X_j \quad (1)$$

Where Y is the predicted responses used as dependent variables, X_i (i = 1, 2 and 3) are the independent variables, and a_0 and a_{ij} (i = 1, 2, 3; j = i, ..., 3) are the model coefficient parameters.

All water extracts were prepared from open air and DIC-assisted Swell- Dried banana peels powders at 5%.

Only the textured banana peels powders underwent a one-night maceration in distilled water at 4°C.

All measurements were made in triplicate, and the results were reported as mean standard deviation. Except for the pivot points matching the experiments run N° 4, 7, 10, 13, 16 and 19 in Table 5, nothing else was repeated.

Variables		Coded	level		
	-1.68	-1	0	1	1.68
X ₁	40	52	70	88	100
X2	60	122	212	302	363
X ₃	10	20	35	50	60
		1 . /	× ¥7	D	

Table 2. Central Composite Design independent variables and their levels (Case of open air)

 X_1 = Extraction Temperature T (°C); X_2 = Particle size (µm); X_3 = Extraction time t (min)

Variables		Coded level						
	-1.68	-1	0	1	1.68			
X ₁	20	60,54	120	179,46	220			
X2	0.3	0,36	0.45	0,54	0.6			
X ₃	1	2,22	4	5,78	7			

Table 3. Central Composite Design independent variables and their levels (case of DIC)

X₁= Treatment time t (s); X₂= Steam Pressure P (MPa); X₃= Number of cycles (-)

Table 4. Central composite design and observed response for nutraceutical compounds and the % of DPPH inhibition of banana peel dried in open air

-	DPPH inhibition of banana peel dried in open air													
	Codeo	d variable	levels	Unco	oded var	iable	Responses							
					levels									
Run	X_1	X_2	X3	X_1	X_2	X3	TFC (mg QE/g	TCC (mg/g d.b)	% of DDPH					
							d.b)		inhibition (Water					
									extract)					
1(C)*	0	0	0	70	212	35	2.57±0.03	0.398 ± 0.005	75.14±0.417					
2	-1	-1	-1	52	122	20	4.00±0.12	0.632±0.010	67.23±0.11					
3	+1	-1	-1	88	122	20	3.5±0.27	0.446±0.031	72.05±0.298					
4	-1	+1	-1	52	302	20	3.69±0.08	0.530 ± 0.005	63.23±0.468					
5	+1	+1	-1	88	302	20	3.55±0.02	0.365±0.011	76.35±0.192					
6	-1	-1	+1	52	122	50	3.80±0.10	0.524±0.018	64.7±0.358					
7	+1	-1	+1	88	122	50	3.38±0.08	0.400±0.019	70.45±0.298					
8	-1	+1	+1	52	302	50	3.16±0.08	0.422 ± 0.005	69.11±0.063					
9	+1	+1	+1	88	302	50	2.93±0.19	0.302 ± 0.057	67.05±0.298					
10(C)*	0	0	0	70	212	35	2.36±0.01	0.414±0.017	64.7±0.358					
11(C)*	0	0	0	70	212	35	2.52±0.02	0.428±0.013	72.23±0.11					
12	-1.68	0	0	40	212	35	2.81±0.10	0.482 ± 0.006	62.57±0.415					
13	+1.68	0	0	100	212	35	2.01±0.02	0.430±0.039	61.6±0.032					
14	0	-1.68	0	70	60	35	3.33±0.16	0.299±0.021	67.35±0.192					
15	0	+1.68	0	70	363	35	5.13±0.32	0.356 ± 0.007	75.58±0.405					
16	0	0	-1.68	70	212	10	2.76±0.11	0.487 ± 0.037	68.23±0.468					
17	0	0	+1.68	70	212	60	2.06±0.01	0.377±0.044	81.03±0.308					
18(C)*	0	0	0	70	212	35	2.61±0.03	0.474 ± 0.041	64.56±0.421					

 X_1 : Extraction Temperature T (°C); X_2 : Particle size Ps (μ m); X_3 : Extraction Time t (min); TFC: Total Flavonoids Content (mg QE/g d.b); TCC: Total Carotenoids Content (mg/g d.b); % of DPPH inhibition (water extract); (C)*: Central point

Table 5. Central composite design and observed response for nutraceutical compounds and % of
DPPH inhibition from Swell-dried textured banana peels by DIC

	Code	ed variat	ole	Uncoded variable levels			Responses			
	levels									
Run	X_1	X_2	X3	X_1	X_2	X3	TFC (mg	TCC	% of DDPH inhibition	
							QE/g d.b)	(mg/g d.b)	(Water extract)	
1(C)*	0	0	0	120.00	0.45	4.00	1.42 ± 0.071	0.43 ± 0.004	66.11 ± 0.685	
2	+1.68	0	0	220.00	0.45	4.00	2.3 ± 0.045	0.78 ± 0.007	71.34 ± 6.219	
3	0	0	+1.68	120.00	0.45	7.00	1.79 ± 0.105	0.59 ± 0.004	68.20 ± 0.117	
4(C)*	0	0	0	120.00	0.45	4.00	1.42 ± 0.071	0.43 ± 0.004	66.11 ± 0.685	
5	0	+1.68	0	120.00	0.60	4.00	1.11 ± 0.17	0.8 ± 0.007	67.46 ± 0.462	
6	0	-1.68	0	120.00	0.30	4.00	1.67 ± 0.032	1.23 ± 0.063	66.41 ± 0.491	
7(C)*	0	0	0	120.00	0.45	4.00	1.42 ± 0.071	0.43 ± 0.004	66.11 ± 0.685	
8	+1	-1	+1	179.46	0.36	5.78	1.74 ± 0.126	0.46 ± 0.008	62.23 ± 0.498	
9	-1	-1	-1	60.54	0.36	2.22	1.26 ± 0.084	0.42 ± 0.008	62.38 ± 0.534	
10(C)*	0	0	0	120.00	0.45	4.00	1.42 ± 0.071	0.43 ± 0.004	66.11 ± 0.685	
11	+1	+1	-1	179.46	0.54	2.22	1.19 ± 0.21	0.99 ± 0.01	66.56 ± 0.471	
12	-1	+1	+1	60.54	0.54	5.78	2.01 ± 0.155	1.06 ± 0.004	70.29 ± 0.222	
13(C)*	0	0	0	120.00	0.45	4.00	1.42 ± 0.071	0.43 ± 0.004	66.11 ± 0.685	
14	-1	+1	-1	60.54	0.54	2.22	2.32 ± 0.182	1.19 ± 0.005	69.55 ± 0.448	
15	-1	-1	+1	60.54	0.36	5.78	2.07 ± 0.195	0.42 ± 0.004	74.77 ± 0.249	

16(C)*	0	0	0	120.00	0.45	4.00	1.42 ± 0.071	0.43 ± 0.004	66.11±0.685
17	-1.68	0	0	20.00	0.45	4.00	3.09 ± 0.371	0.44 ± 0.004	67.76 ± 0.249
18	+1	+1	+1	179.46	0.54	5.78	2.04 ± 0.351	0.54 ± 0.006	68.65 ± 0.241
19(C)*	0	0	0	120.00	0.45	4.00	1.42 ± 0.071	0.43 ± 0.004	66.11± 0.685
20	+1	-1	-1	179.46	0.36	2.22	3.65 ± 0.138	0.49 ± 0.003	69.10 ± 0.132
21	0	0	-1.68	120.00	0.45	1.00	3 ± 0.184	0.48 ± 0.003	69.25 ± 0.209
22(C)*	0	0	0	120.00	0.45	4.00	2.89 ± 0.032	0.64 ± 0.003	70.14 ± 1.053

 X_1 : Treatment time t (s); X_2 : Steam pressure P (MPa); X_3 : Number of cycles (-); TFC: Total Flavonoids Content (mg QE/g d.b); TCC: Total Carotenoids Content (mg/g d.b); % of DPPH inhibition (water extract); (C)*: Central point

2.2.3. Physicochemical analysis

• Hexane extraction was used to extract the oil from banana peels. The fatty acid profile (Table 6) was determined using CHROMPACK CP 9002 gas phase chromatography. The methyl esters were produced by esterification using the ISO 5509 technique (ISO, 2000).

• An Atomizer (VARIAN AA 240, Australia) was used to determine the mineral composition (Table 7) of banana peels. This measurement is based on dissolving 1g of ashes in 5mL of HCL acid (0.5N) (Adrian, et *al.* 1995).

• The mucilage was extracted from banana peel powder according to the method described by Dick et *al.* (2019). 30 g of banana peel powder were mixed with 100mL of distilled water and heated for 54 to 96 minutes at 50°C. The mucilage was lyophilized after being precipitated with 35mL of ethanol.

• A colorometric method was used to determine the Total Flavonoids Content (TFC) (Bahorun et *al.* 1996). 1mL of the water extract was mixed with 1mL of a 2% AlCl3 solution. After a 10 minutes incubation period, the absorbance was measured using a spectrophotometer at 430nm. The TFC in different banana peel water extracts was estimated using a regression equation that used Quercetin as a standard and was estimated in milligrams of Quercetin Equivalent per gram dry basis (mg QE/g d.b).

The Total Carotenoids Contents TCC was extracted using the Sass-Kiss et al. (2005) method. 20 mL of a solvent mixture of hexaneacetone-ethanol (2V: 1V: 1V) were added to 0.5g of banana peel powder. After 30 minutes of agitation, the upper phase was recovered. For a second extraction, 10 mL of hexane were added. The absorbance was measured using a spectrophotometer at 450nm. Carotenoid concentrations were estimated by referring to the regression equation with carotene as a standard and were expressed in milligrams per gram dry basis (mg /g d.b).

• The DPPH free radical scavenging method was employed to assess the antioxidant activity of aqueous extracts of banana peels dried in the open air and DIC-assisted Swell–drying (Kroyer, and Hegedus, 2001). A volume of 0.1mL of extract was added to 3.9mL of DPPH (60mM). After 30 minutes of incubation in the dark, the absorbance was measured at 517 nm. The percentage of DPPH radical inhibition is expressed as follows:

% of DPPH radical inhibition = $\frac{(\text{OD517 of Reference} - \text{OD517 of water extract})}{\text{OD517 of Reference}} \times 100$

(2)

Fatty Acids	Names	Content (%)
C12 :0	Lauric A.	1.32
C16 :0	Palmitic A.	28.32
C16 :1 ω 7	Palmitoleic A.	1.53
C18 :0	Stearic A.	5.54
C18 : 1 ω 9	Oleic A.	3.93
C18 : 2 ω 6	Linoleic A.	19.35
C18 : 3 ω 3	Linolenic A.	22.13
C22 :0	Behenic A.	3.13
C14 : 0	Myristic A.	1.50
C24: 0	Lignoceric	2.81

Table 6. Fatty acids (%) of Musa acuminate peel powder

Table 7. Mineral composition of Musa acuminate peel powder

Minerals	Content (g/Kg d.b)
Ca	30.254
Mg	25.836
Na	54.701
Κ	24.178
Fe	0.441
Cu	0.019
Zn	0.018
Cr	-
Cd	-
Pb	-

2.2.4. Statistical analysis

Using Statgraphics Centurion 18 software, the Response Surface Methodology (RSM) was utilized to define the relationship between independent variables and to deduce the optimum extraction conditions of bioactive substances from banana peel powders. A quadratic model to predict the response (TFC, TCC and percentage of the DPPH radical inhibition) was determined from all the experiments, and the regression coefficients for the linear, quadratic and interaction factor were calculated and statistically examined using variance analysis (ANOVA). The multiresponse optimization was defined using the desirability approach (Response Optimizer function in Statistica). The desirability function is a multi-criteria approach for assessing the overall desirability that considers each response's desirable value or the best accepted.

The Surface Responses were generated using the polynomial equations regressions coefficients. These were utilized to analyse the variables under investigation as a function and to present the most favourable acceptance.

To highlight the interaction effects of independent variables of each drying process, one variable remained constant at its centre level while the other two variables changed within the experimental range.

3.Results and discussions

3.1. Physicochemical analysis

Musa acuminata peel dried in the open air studied here contains oil with a yield of $10\pm0.5\%$. This oil is rich in unsaturated fatty acids such as palmitic acid (28.32%), linolenic acid (22.13%), linoleic acid (19.35%) and oleic acid (3.39%) (Table 6). These fatty acids are beneficial to people's health and are thought to help in the prevention of cancer, atherosclerosis and obesity (Kaleem, 2013).

High levels of macro-metals (30.02; 5.47; 2.583 g/Kg d.b) for Ca, Na and Mg respectively and low levels of trace metals (Fe, K) were found in *Musa acuminata* peel (Table 7). Heavy metals like Cr and Cd are not present.

Banana peel, which is rich in minerals, could be used as a powder to correct mineral deficiencies.

The size of the particle determines how much mucilage can be produced from banana peels powder $21.25\pm12.37\%$ (for a particle size $\Phi=363\mu$ m); $17.25\pm3.181\%$ ($\Phi=208\mu$ m); $14\pm5.656\%$ ($\Phi=69\mu$ m).

The amount of mucilage is higher than that of flaxseed, which Mazza and Biliaderis, (1989), and Benahmed Djilali et *al.* (2022) reported to yield, respectively, 7.08 and 9.4%.

Additional factors, such as the temperature and extraction time, have also an impact on the mucilage content (Mazza, and Biliaderis, 1989).

The mucilage of banana peels was verified using the FT-IR method (Fig.1). A strong band is visible at 3438 cm-1 (-OH stretching) and 2935 cm-1 (-CH stretching) is observed (Sawut et *al.* 2014).

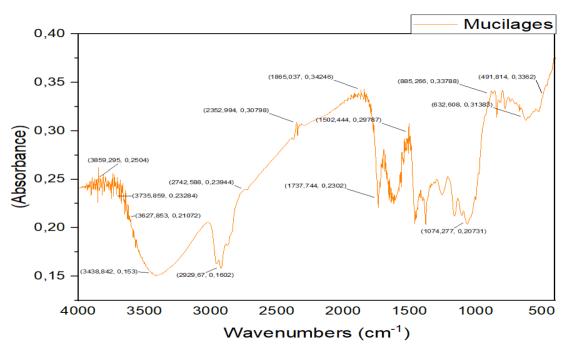


Figure 1. FT-IR spectrum of banana peel mucilage

The vibrational peaks at 1215 and 1074 cm-1 (CO, ether) and (C-C of pyrenoids), respectively, indicate the presence of polysaccharides (mucilage) in banana peel (Pereira et *al.* 2017).

The peaks at 1750 and 1733 cm-1 are caused by stretching vibrations of the C=O group (Kpodo et *al.* 2017).

The carboxyl group COOH of Galacturonic Acid (GalA) is responsible for the peaks between 1650 and 1733 cm-1 (Nejatzadeh-

Barandozi, and Enferadi, 2012; Kpodo et *al.* 2017).

In the food industry, this polysaccharide can be used for stabilizing beverages and adding texture to dairy desserts, among other uses (Qin et *al.* 2005).

3.2. Fitting the models

3.2.1. Case of open air drying

The following empirical models for the Total Flavonoids Content, Total Carotenoids Content and % of the DPPH inhibition from open air banana peels powder and their regression (Table 8).

Total Flavonoids Content $\left(mg\frac{QE}{g}d.b\right)$

Where X_1 : Extraction Temperature T (°C); X_2 : Particle size Ps (μ m); X_3 : Extraction Time t (min).

(3)

(4)

(5)

 $= 8.661 - 0.047X1 - 0.036X2 - 0.016X3 + 0.0002X1^2 + 0.00004X1X2$

 $-0.000005X1X3 + 0.00009X2^2 - 0.00008X2X3 + 0.0003X3^2$

Total Carotenoids Content(mg QE/g d. b)

 $= 1.136 - 0.014X1 + 0.001X2 - 0.009X3 + 0.00006X1^2$

 $+ 0.000002X1X2 + 0.00005X1X3 - 0.000003X2^2 - 0.000002X2X3$

 $+ 0.00005X3^{2}$

% of DPPH radical inhibition (water extract)

 $= 22.172 + 1.358X1 - 0.041X2 - 0.131X3 - 0.007X1^2$

 $+ 0.000038X1X2 - 0.0066X1X3 + 0.00011X2^{2} + 0.000066X2X3$

 $+ 0.0092X3^{2}$

The fitted models for TFC and TCC (Eq.2 and Eq.3) using the open-air drying process were satisfactory, with significant regressions responsible for residual values and satisfactory determination coefficients R^2 (76 and 68%), which represents acceptable equation fitting. However, the percentage of DPPH inhibition, presents a low value of R^2 (54%) (Table 8).

Statistical analysis of the experimental design shows that the Total Flavonoids Content extract from open air banana peels powder is only affected by DIC operating parameters (particle size); the quadratic second-order effect was significant (P<.05) as observed in Fig.2.

Fig. 3 and Eq. 3 allow noting the extraction temperature T has an impact on the quantity of

Total Carotenoids extracted from open air banana peels powder (a negative linear firstorder effect was significant (P < .05)). When the extraction temperature decreases the TCC increases.

Here too, statistical analysis showed that the percentage of DPPH inhibition was not affected by any of the studied parameters (Temperature, Particle size and time), and the other terms from eq.4 are not statistically significant (P>05) (Fig.4).

The absence of interaction effects for TFC, TCC, and the percentage of DPPH inhibition suggests that there is no synergistic interaction between the components being studied.

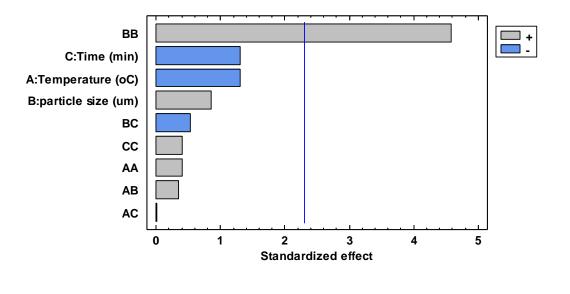
	TFC		TCC		% of DPPH inhibition (Water extract)	
Regression	Values	<i>P</i> -value	Values	P-value	Values	<i>P</i> -value
coefficients						
a 0	8,66179		1,13642		22,1717	
a 1	-0,0472928	0,2291	-0,0139118	0,0247	1,35838	0,3235
a ₂	-0,0356211	0,4181	0,00101351	0,2785	-0,0407254	0,4486
a3	-0,0162992	0,2269	-0,00940816	0,0736	-0,130885	0,4893

Table 8 Regression coefficients of the second-order polynomial and their significance (open air)

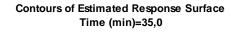
a11	0,000197904	0,6917	6,41098E-05	0,3089	-0,0075248	0,1363
a 22	8,6661E-05	0,0018	-3,13178E-06	0,2143	0,00011356	0,543
a 33	0,000284981	0,6917	0,0000539181	0,5434	0,00923629	0,1957
a ₁₂	4,24383E-05	0,7315	1,92901E-06	0,8984	3,78086E-05	0,9741
a ₂₃	-7,68519E-05	0,6063	-1,57407E-06	0,9308	6,57407E-05	0,9624
a ₁₃	-4,62963E-06	0,995	4,9537E-05	0,5883	-0,00659722	0,358
\mathbb{R}^2	76,43 percent		68,97 percent		54,62 percent	

Α

Standardized Pareto Chart for TFC (mg QE/g d.b)



В



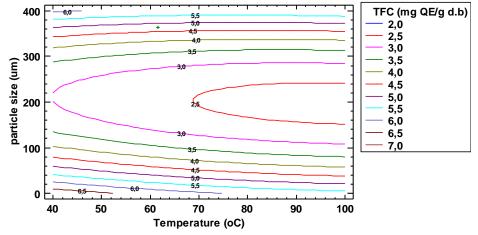
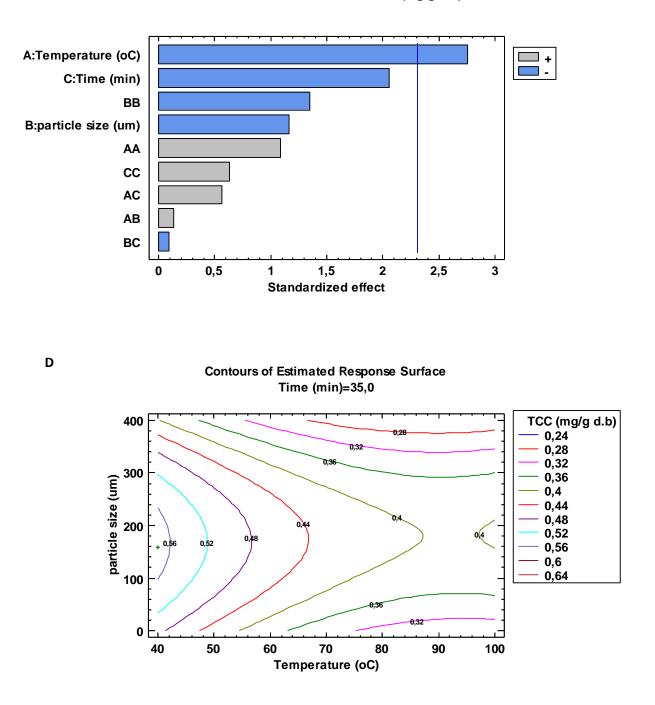
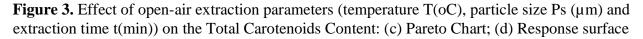


Figure 2. Effect of open-air extraction parameters (temperature (oC), particle size (μm) and extraction time (min)) on the Total Flavonoids Content: (a) Pareto Chart; (b) Response surface

С

Standardized Pareto Chart for TCC (mg/g d.b)





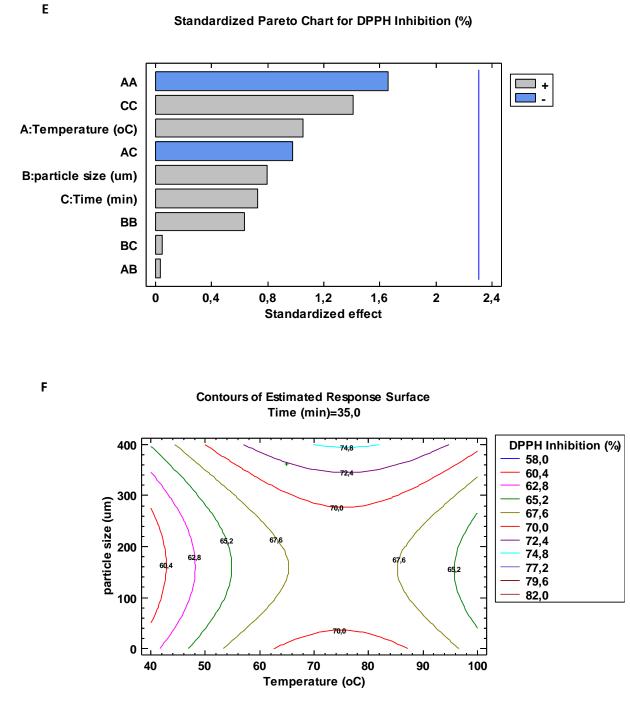


Figure 4. Effect of open-air extraction parameters (temperature T(oC), particle size Ps (μ m) and extraction time t(min)) on the Total Carotenoids Content: (e) Pareto Chart; (f) Response surface

3.2.2. Case DIC assisted Swell-drying

The following responses form of a secondorder polynomial for TFC, TCC and % of the DPPH inhibition from Swell-Dried banana peels powders and their regression (Table 9). Where X_1 : Treatment time t (s); X_2 : Steam pressure P (MPa); X_3 : Number of cycles (-).

Total Flavonoids Content $\left(mg \frac{QE}{g} d. b \right)$ (6) = 0,258343 + 0,0161853X1 + 14,0994X2 - 1,08437X3 + 0,0000997754X1² - 0,0738125X1X2 - 0,00184242X1X3 - 13,4117X2² + 1,27965X2X3 + 0,0778587X3² Total Carotenoids Content $\left(mg \frac{QE}{g} d. b \right)$ (7) = 3,0694 + 0,00720274X1 - 15,9562X2 + 0,175842X3 + 0,0000130335X1² - 0,0193875X1X2 - 0,000413364X1X3 + 23,3667X2² - 0,429151X2X3 + 0,00617456X3² % inhibition of DPPH radical of water extract (8) = 57,4982 + 0,00424404X1 + 0,00424404X2 + 2,16904X3 + 0,000260012X1² + 0,0277965X1X2 - 0,0211525X1X3 - 0,650717X2² - 2,09894X2X3 + 0,198078X3²

The DIC technique was used to successfully fit the TFC and TCC (Eq.5 and Eq.6). The resulting models were suitable with acceptable determination coefficients R^2 , with similar values (~ 56%), indicating satisfactory

equation fitting and appropriate adjustment models, except for the percentage of DPPH inhibition, which had a low value of R^2 (42.55%) (Table 9).

	TFC		TCC		% of DPPH inhibition (Water extract)	
Regression coefficients						
	Values	<i>P</i> -value	Values	<i>P</i> -value	Values	<i>P</i> -value
a 0	0,258343		3,0694		57,4982	
a ₁	0,0161853	0,874	0,00720274	0,9658	0,00424404	0,6759
a 2	14,0994	0,3739	-15,9562	0,1718	12,4278	0,4358
a ₃	-1,08437	0,2764	0,175842	0,6346	2,16904	0,5362
a 11	9,97754E-05	0,0436	1,3034E-05	0,4568	0,00026001	0,2202
a ₂₂	-13,4117	0,5007	23,3667	0,0082	-0,650717	0,9942
a 33	0,0778587	0,1408	0,00617456	0,7497	0,198078	0,3946
a ₁₂	-0,0738125	0,0947	-0,0193875	0,2372	0,0277965	0,883
a 23	1,27965	0,365	-0,429151	0,4258	-2,09894	0,7398
a 13	-0,00184242	0,388	-0,00041336	0,6094	-0,0211525	0,0429
\mathbb{R}^2	55,84 percent		56,07 percent	•	42,55 percent	•

Table 9 Regression coefficients of the second-order polynomial and their significance (DIC)

According to the study of the variance, Fig. 5 and Eq.5 allow noting that DIC treatment time t is only significant for the increase of Total Flavonoids Content (a positive quadratic effect was significant (P < .05)).

Statistical analysis of the experimental design showed the steam pressure P to be the only significant DIC operating condition affecting the increase of Total Carotenoids recovered from DIC swell-dried banana peels powder (a positive quadratic second-order effect was significant (P < .05)) (Fig.6).

Statistical analysis of the experimental design allowed obtaining Pareto Chart and Response surface (Fig.7), as well as the empirical model (Eq.7), which show that treatment time t and number of cycles are the main significant operating parameters. (a negative linear effect was significant (P<.05)). When the treatment time t and the Number of

cycles increase, the percentage of DPPH inhibition decreases.

3.3.Optimization of extraction conditions

3.3.1. Case of open air

The optimal values for the variables used in the open air drying of banana peels, with maximum desirability were found to be as follows: temperature, 60.47°C; extraction time, 10min; and particle size, 348.648µm. The predicted values are satisfactory because the optimal desirability d=0.7.

The values for the responses at the optimized point were as follows (Fig.8):

Total Flavonoids Content= 5.13 (mg QE/g d.b) with a desirability d=1;

Total Carotenoids Content = 0.48 (mg/g d.b) with a desirability d=0.54;

% of DPPH inhibition=73.05% with a desirability d=0.63.

Standardized Pareto Chart for TFC (mg QE/g d.b)

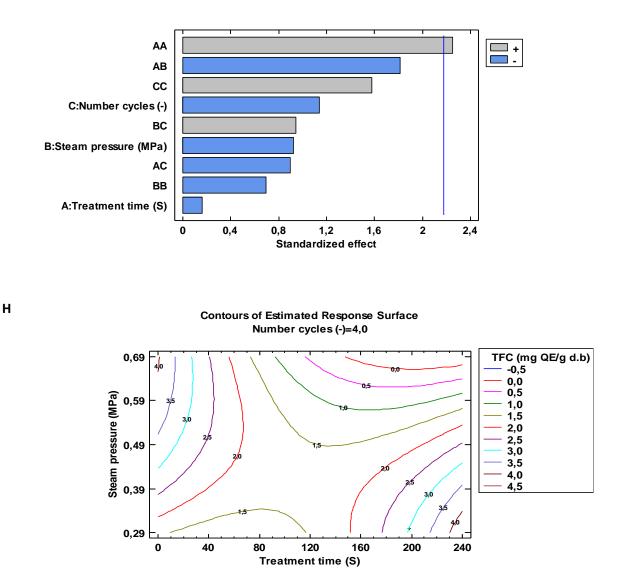


Figure 5. Effect of Swell-Drying with DIC parameters (Treatment time t(s), steam Pressure P (Mpa) and Number of Cycles)) on the Total Flavonoids Content: (g) Pareto Chart; (h) Response surface

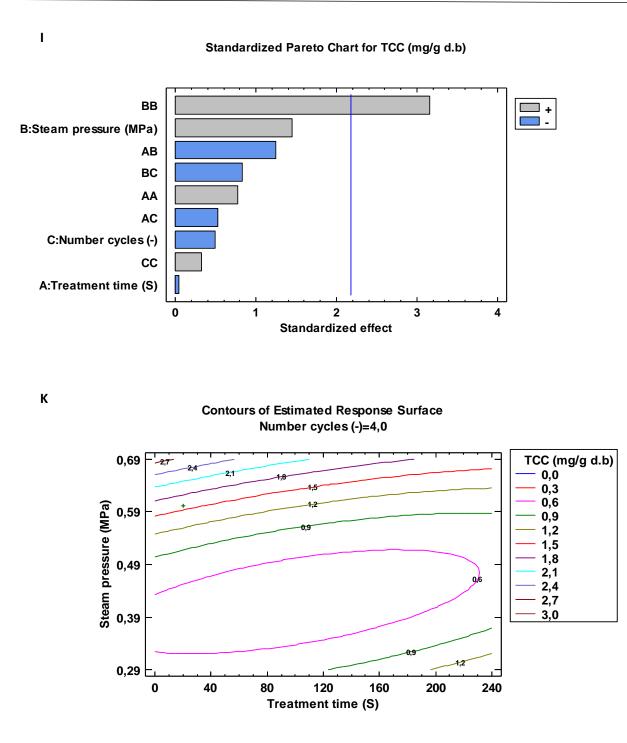


Figure 6. Effect Swell-Drying with DIC parameters (Treatment time t(s), steam Pressure P(Mpa) and Number of Cycles)) on the Total Carotenoids Content: (i) Pareto Chart; (k) Response surface

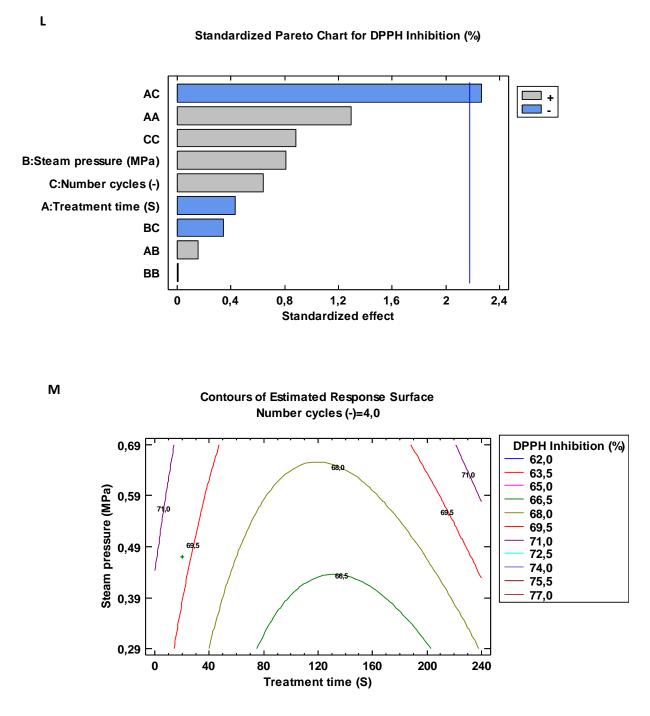


Figure 7. Effect Swell-Drying with DIC parameters (Treatment time t(s), steam Pressure P(Mpa) and Number of Cycles) on the % of the DPPH inhibition: (l) Pareto Chart; (m) Response surface

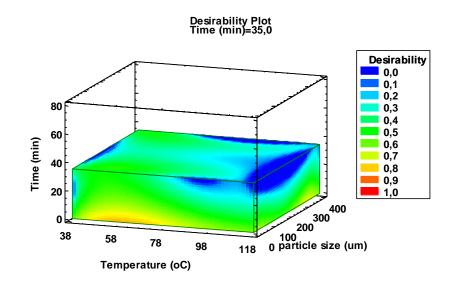


Figure 8. Response surface of the effect of open-air extraction parameters (temperature (oC), particle size (µm) and extraction time (min)) on desirability

3.3.2. Case of Swell- drying by DIC

The optimized DIC operating parameters with maximum desirability were found to be as follows: treatment time, 24.46 s; steam pressure, 0.59MPa; number of cycles, 6.38.

Fig.9 allowed obtaining the following response with the highest optimized desirability (d=1):

TFC=4.07 (mg QE/g d.b), TCC= 1.37 (mg /g d.b) DPPH inhibition=75.97 % Hence, the predicted values are perfect (d=1).

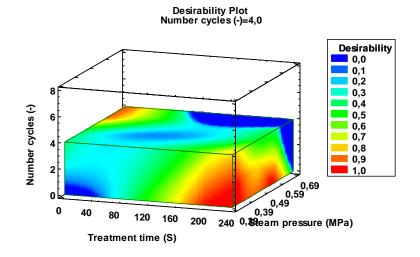


Figure 9. Effect of DIC-assisted Swell-Drying parameters (Treatment time t (s), steam Pressure P (Mpa) and Number of Cycles Nc)) on the Desirability

3.4. Discussions

The Response optimizer Function was used to optimize the operating conditions of two drying processes (open air and DIC assisted Swell Drying) in order obtain a maximum of nutraceutical components (flavonoids and carotenoids) from banana peels.

In this study, various quantitative findings on the biochemical properties were investigated. This concerns minerals, fatty acids, mucilage, flavonoids, and carotenoids.

The optimized extraction parameters for open-air dried banana peel were found at 60.47° C, extraction time (10min) and particle size (348.648µm). Additionally, the thermal treatment time (24.46s), saturated steam pressure (0.59MPa) and cycles Number 6.38 were the optimized operating DIC texturing parameters that were found. Under the selected optimum conditions, the suitability of the models to predict the responses of the Total Flavonoids Content and Total Carotenoids Content and % of DPPH inhibition was evaluated.

The richness of banana peels in nutraceuticals compounds (flavonoids and carotenoids) was linked to the drying processes and the operating conditions. The highest optimized concentration of Total Flavonoids Content (5.13 mg QE/g d.b) was observed from open-air dried banana peels and correlated positively with the particle size.

However, the TFC released from DIC Swell-dried banana peels was correlated positively with the heat treatment time with DIC at the optimum value (4.07 mg QE/g d.b).

In plant tissues, phenolic compounds appear as glucosides and have a lot of hydroxyl groups. This produces a large variety of these molecules with various properties, including solubility (Muzolf-Panek, and Gliszczynska-Swiglo, 2022).

Using water as an extraction solvent, in order to hydrate the particle, which intensifies mass transfer by diffusion (Ghitesci et *al.* 2015).

Our findings are comparable to those found by Someya et *al.* (2002), who showed that banana peels had a high level of total phenolic content (907mg/100g). On the other hand, banana peels heat-dried by microwave irradiation have a higher phenolic component level than freeze-dried banana peels (Vu et *al*. 2018).

Conversely, increasing temperature resulted in a loss of sensitive compounds and decreased the antioxidant activity. According to Passo Tsamo et *al.* (2015), boiling banana peels decreased the amount of flavonoids while increased the ferulic acid.

According to earlier studies, the use of other extraction procedures was responsible for the loss of some phenolic compounds (Gonzalez-Montelongo, 2010; Toh et *al.* 2016; Hernandez-Carranza et *al.* 2016).

The variation of phenolic compounds depends on the variety, cultivation condition and the maturity of fruit.

Additionally, it can be said that the heat treatment with DIC is an effective process for releasing bound phenolic compounds from their phenolic acids. This has attracted a lot of interest due to their antioxidant qualities.

When using DIC-texturing, biological composition is preserved thanks to the expanded granule powder with a high level of interaction (porosity and capillarity) (Benahmed Djilali et *al.*, 2016; Benseddik et *al.*, 2022).

The level of carotenoids was maximised at 1.72 mg/g d.b by using Swell-drying with DIC.

Our findings are comparable to those of Nguyen et *al.* (2014), who showed that carrots subjected to Swell-Drying assisted by DIC, which maximised porosity for the steam pressure (P=0.5MPa) and thermal holding time (~ 36 s). Furthermore, the optimal effectiveness of an extract cannot result from one active compound but rather form the combined synergistic action of various constituents.

The effects of temperature, extraction time, and methanol concentration on the TFC from *Nigella sativa* seeds were examined in the study by Muzolf-Pan and Gliszczynska-Swiglo (2022). TFC increased with an increase in temperature and extraction time up to T = 670C, t = 208 min, and methanol concentration = 50%, allowing the maximum predicted value of TFC =7.68 mg QE/g.

4. Conclusions

Some operational parameters of the two drying processes (open air and Swell-Drying with DIC were optimized to maximise the availability of Total Flavonoids and carotenoids using single- and multi-response desirability functions.

Texturing banana peel by DIC promotes high levels of carotenoids and maximised the antioxidant activity

Banana peel water extract could be a good source of bioactive substances, which could be further used as a natural antioxidant in various industrial settings.

However, for the sake of this study it would be interesting to apply the optimum operational parameters resulting from the open air to the second drying process.

Future study will focus on identifying the phenolic components of the obtained powders, optimizing additional parameters, and using those components to elaborate medicinal forms.

5.References

- Adrian, J., Potus, J., Frangne, R. (1995). Food Science of A to Z. Ed Tech and Doc. Lavoisier, Paris
- Bahorun, T., Gressier, B., Trotin, F., Brunet, C., Dine, T., Vasseur, J., Gazin, J.C., Pinkas M., Luyckx, M., and Gazin, M. (1996).
 Oxygen species scavenging activity of phenolic extracts from hawthorn fresh plant organs and pharmaceutical preparations. *Arzneimttel-forschung and Drug Research*, 46 (11), 1086.
- Benahmed Djilali, A., Benseddik, A., Metahri,
 M.S., Lahouazi, D., Simoud, D., Nabiev,
 M., Allaf, K. (2022). Elaboration of new
 functional dairy dessert based on flaxseed
 powder. 1 st International online
 Conference on Agriculture-Advances in
 Agricultural Science and Technology,
 session Seed Sciences and Technology.
 Chemistry. Processes.

- Benahmed Djilali, A., Nabiev, M., Gelicus, A., Benamara, S., & Allaf, K. (2016).
 Evaluation of Physical-Chemical, Pharmacodynamic and Pharmacological Attributes of Hot Air Dried and Swell Dried Jujube Powders. *Journal of Food Process Engineering*, 40, 2, e12364.
- Benseddik, A., Benahmed-Djilali, A., Azzi, A.,
 Zidoune, M.N., Bensaha, H., Lalmi, D., and
 Allaf, K. (2022). Effect of Drying Processes
 on the Final Quality of Potimarron
 Pumpkin (*Cucurbita maxima*) Powders.
 Journal Dispersion Science and
 Technology, 43(1), 136-146.
- Dantas, T.L., Alonso Buriti, F.C., Florentin, E.R. (2021). Okra (*Abelmoschus esculentus* L.) as a functional food source with potential mucilage and bioactive compounds that has technological applications and health benefits. *Plants*, 10(8), 1683.
- Dick, M., Dal Magro, L., Rodrigues, R.C., De Rios, A., Flores, SH. (2019). **O**. Valorization of *Opuntia* monacantha (Willd). Haw. Cladodes to obtain mucilage with hydrocolloid features physicochemical and functional performance. International Journal of Biological Macromolucles, 123, 900-909.
- FAO statistical database (2022).
- Gemede, H.F., Retta, N., Haki, G.D., Woldegiorgis, A.Z., Beyene, F. (2015). Nutritional quality and health benefits of Okra (Abelmoschus esculentus). International Journal of Nutrition and Food Sciences, 4(2), 208-215.
- Ghitesci, RE., Volf, I., Carausu, C., Beuhlmann, A.M., Gilca, I.A., Popa, V.I. (2015). Optimization of ultrasound-assisted extracyion of polyphenols from spruce wood bark. *Ultrasonics Sonochemistry*, 22, 535-541.
- Gonzalez-Montelongo, R., Lobo, G., Gonzalez, M. (2010). Antioxidant activity in banana peel extracts: testing extraction conditions and related bioactive compounds. *Food Chemistry*, 119, (3), 1030-1039.

- Happi Emaga, T., Herinavalona Andrianaivo, R., Wathelet, B., Tchango, J., Paquot M. (2007). Effects of the stage of maturation and varieties on the chemical composition of banana and plantain peels. *Food Chemistry*, 103(2), 590-600.
- Havsteen, B. (1983). Flavonoids, a class of natural products of high pharmacological potency. *Biochemical and Pharmacology*, 32(7), 1141-1148.
- Hernandez-Carranza, P., Avila-Sosa, R., Guerrero–Beltran, J.A. Navarro-Cruz, A.R., Corona-Jimenez, E., Ochoa-Velasco, C.E. (2016). Optimization of Antioxidant compounds extraction from fruit by products: Apple pomace, orange and banana peel. *Journal Food Processing and Preservation*. 40(1), 103-115.
- Iman, M.Z & Akter, S. (2011). Musa paradisiaca L. and *Musa sapientum* L. A phytochemical and pharmacological review. *Journal of Applied Pharmaceutical Science*, 1, 14-20.
- ISO 5509 technique. (2000).
- Kaleem, M. (2013). Effects of products oxidation of linoleic acid on ruminal biohydrogenation. Doctoral Thesis from Toulous Univrsity, France.
- Kpodo, F.M., Agbenorhevi, J.K., Alba, K., Bingham, R.J., Oduro, I.N., Morris, G.A., Kontogiorgos, V. (2017). Kontogiorgos. Pectin isolation and characterization from six okra genotypes. *Food Hydrocolloids*, 72, 323-330.
- Kroyer, G., & Hegedus, N. (2001). Evaluation of bioactive properties of pollen extracts as functional dietary food supplement. *Innovative Food Science and Emerging Technologies*, 2(3) 171-174.
- Mazza, G., & Biliaderis C.G. (1989). Functional properties of flax seed mucilage. *Journal of Food Science*, 54(5), 1302-1305.
- Muzolf-Panek, M., & Gliszczynska-Swiglo. (2022). Extraction optimization for the antioxudants from *Nigella sativa* seeds using response surface methodology. *Journal of Food Measurement and Characterization*.

- Nejatzadeh-Barandozi, F., Enferadi, S.T. (2012). FT-IR study of the polysaccharides isolated from the skin juice, gel juice, and flower of aloe vera tissues affected by fertilizer treatment. *Organic and Medicinal Chemistry Letters*, 2, 33.
- Nguyen, T.H., Lanoiselle, J.L, Allaf, K. (2014). Effect of Instant Controlld pressure Drop DIC treatment on the properties of dried carrot. 19 th International drying symposium IDS France.
- Passo Tsamo, CV., Herent, M.F., Tomekpe, K., Happi Emaga, T., Quetin-Leclercq, J., Rogez, H., Larondelle, Y., Andre, C.M. (2015). Effect of boiling on phenolic profiles determined using HPLC/ESI-LTQ-Orbitrap-MS, physico-chemical parameters of six plantain banana cultivars (*Musa sp*). *Journal of Food Composition and Analysis*, 44, 158-169.
- Pereira, G.A., Moulina, G., Arruda, HS., Pastore, G.M. (2017). Optimizing the Homogenizer- Assisted Extraction (HAE) of Total Phenolic Compounds from Banana Peel. *Journal of Food Process Engineering*. 40(3), e12438.
- Qin, L., Xu, SY., Zhang, W.B. (2005). Effect of enzymatic hydrolysis on the yield of cloudy carrot juice and the effects of hydrocolloids on color and cloud stability during ambient storage. *Journal of Science Food and Agriculture*, 85(3), 505-512.
- Sass-Kiss, A., Kiss, J., Milotay, P., Kerek, M.M, Toth-Markus, M. (2005). Differences in anthocyanin and carotenoid content of fruits and vegetables. *Food Research International*, 38(8-9), 1023-1029.
- Sawut, A., Yimit, M., Sun, W., & Nurulla, L. (2014). Photopolymerisation and characterization of maleylatedcellulose-gpoly (acrylic acid) superabsorbent polymer. *Carbohydrate Polymers*, 101, 231-239.
- Someya, S., Yoshiki, Y., and Okubo, K. (2002). Antioxydant compounds from banana (*Musa Cavendish*). *Food Chemistry*, 26(3), 2561.
- Toh, P.Y., Leong, F.S, Chang, S.K., Khoo, H.E., Yim, H.S. (2016). Optimization of

Extraction parameters on the oxydant properties of banana waste. Acta Scientiarum Polonorum Technologia Alimentaria, 15(1), 65-78.

Vu, H.T., Scarlett, C.J., Vuong, Q.V. (2018). Phenolic compounds within banana peel and their potential: A review. *Journal of Functional Foods*, 40, 238-248.

6. Abbreviations

- db Dry basis (g)
- DIC Instant Controlled Pressure-Drop
- DPPH 2,2-diphenyl-1-picrylhydrazyl
- SD Swell-Drying
- SD Swell-Drying
- TFC Total Flavonoids Content
- TCC Carotenoids Content