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COMPARATIVE ANTIOXIDANT AND PHYTOCHEMICAL ACTIVITY OF RAW AND BOILED TUBER OF *DIOSCOREA BULBIFERA* COLLECTED FROM TRIBAL FOREST OF SUNDARGARH DISTRICT, ODISHA, INDIA

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| Article history: | ABSTRACT |
|--|---|
| Received: 18 September 2021 | In the present study, we have investigated the differences in the composition |
| Accepted: 17 July 2023 | of proximate minerals, vitamins bioactive compounds, and 1,1-diphenyl-2- |
| Keywords: Antioxidant; Ascorbic acid; Bioactive compound; Flavonoid. | picrylhydrazyl (DPPH) scavenging activity between the raw and boiled tubers of <i>Dioscorea bulbifera</i> . The results showed that both the raw and boiled tubers have rich sources of carbohydrates (31.62% and 23.94%), proteins (3.48% and 2.25%), starch (8.6% and 11.67%), and free amino acids (1.45% and 0.59%); but have low-fat content (0.19% and 0.14%). Vitamin profiling of the tubers contained a substantial amount of ascorbic acid, vitamins B1, B2, B3, and B6. Further, the raw and boiled tuber of <i>Dioscorea bulbifera</i> had a very high amount of bioactive compounds like phenolics, flavonoid, diosgenin, tannin, and saponin. Phenolic and flavonoid content positively correlated with free radical scavenging activity of tuber and performed better scavenging activity compared to ascorbic acid and butylated hydroxytoluene (BHT). Thus, the tuber of <i>Dioscorea bulbifera</i> is a better food supplement to meet the calorie requirement of the tribal people and a rich source of antioxidants. |

1.Introduction

Wild tubers *Dioscorea bulbifera*, known as air potato or yam, is primarily found in the tropical, subtropical, and temperate region of the world (Abara, 2011). *Dioscorea bulbifera* produces both underground and aerial tubers. However, underground tubers are very rich in starch hence mostly consumed. The tribal people used it as food as well as medicine (Kumar *et al.*, 2013). The tubers are composed of a high amount of carbohydrates, fibers, and low fats and protein content with a good proportion of different amino acids, making them an excellent dietary source and could be consumed, boiled, steamed, baked, or fried (Osman, 1990). Due to lack of good nutritional information, the broad

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utilization of the yam is very limited; Studies of the nutritional composition of the locally available Dioscorea bulbifera is very important since it may help the tribal people to fulfill their dietarv requirements. Besides excellent nutritional properties, Dioscorea bulbifera reported exhibiting antimicrobial, antioxidant, plasmid curing, analgesic, anti-inflammatory, antihyperglycemic, antihyperlipidemic, antidiabetic, antinociceptive, and antitumor activities (Ghosh et al., 2015). Dioscorea bulbifera is also reported to have good radical scavenging and singlet oxygen quenching ability hence used as potential herbal therapeutic agents for various diseases that occurred due to

oxidative stress (Ghosh *et al.*, 2013). *Dioscorea bulbifera* (*D. bulbifera*) exhibits higher antioxidant capacities with lower IC₅₀ values than the other species (Padhan *et al.*, 2020). Detailed information on the profiling of tubers of *D. bulbifera* from the state of Odisha is very rare. In this regard, the present study is therefore aimed to evaluate the comparative phytochemical and antioxidant properties of the boiled and raw tuber of wild yam *D. bulbifera*.

2. Materials and methods

2.1. Collection and preparation of sample

The tubers specimen and the plant were collected from the forest of Sundargarh, Odisha, and identified at the Centre of Excellence in Natural Products and Therapeutics, Dept. of Biotechnology and Bioinformatics, Sambalpur University. Tubers were washed thoroughly, peeled, and sliced approximately 1-2 mm of thickness. Some of the sliced tubers were boiled and dried in a hot air oven at 80 °C until a constant weight was obtained. In contrast, the other portion was taken fresh, without boiled, and dried in a hot air oven at 80 °C until a constant weight was obtained. After drying, the samples were powdered and sieved through a 1mm sieve. Both raw and boiled tuber powder samples were preserved in an airtight glass bottle for further analysis.

2.2. Comparative Nutritional analysis

Nutritional analyses of both raw and boiled tuber were determined. Moisture, ash, and fat contents were determined using a hot air oven, muffle furnace and Soxhlet apparatus following the standard method described by Ranganna (2007). Total carbohydrate and starch content were estimated using the Anthrone reagent method (Thayumanavan and Sadasivam, 1984). Reducing sugar content of both the raw and boiled tuber samples was estimated by the Dinitrosalicyclic acid method (Sadasivam and Manickam, 2008). The protein content of both the raw and boiled tuber samples was estimated following the standard method of Lowery et al. (1951). The total free amino acid of the tuber samples was determined using Ninhydrin

reagent (Sadasivam and Manickam, 2008). The amino acid profiling of the raw and boiled tuber samples was determined using 5.54 SP 5LAB solutions software after detection through Shimadzu LC-30 AD HPLC (Pal et al., 2016). Vitamins (B1, B2, B3, and B6) in tuber samples were analyzed in HPLC (Shimadzu HPLC and photodiode array detector. Supelcosil LC 17 DB column 250 mm×4.6 mm, 5µm; Sigma, USA) following the method of Perales et al. (2005). The mineral compositions were analyzed using an Inductively Coupled Atomic Adsorption Spectrometer (Perkin-Elmer Optical Emission Spectrometer, Optima 7000 DV) as per the methods of Kalra (1998). The ascorbic acid content was estimated as the standard method described by Sadasivam and Manickam (2008).

2.3. Comparative Bioactive compounds analysis

Total free phenolics and Total flavonoid content were determined in methanol, acetone, and water extract of tuber using modified Folin-Ciocalteau method (Ordon Ez *et al.*, 2006) and the aluminum chloride colorimetric assay (Pallab *et al.*, 2013), respectively. Tannin was estimated by following the methods of Schanderl (1970). The method of Obadoni and Ochuko (2001) was used for the determination of saponin content. Diosgenin was determined following the methods of Uematsu *et al.* (2000).

2.4. DPPH scavenging activity

Tuber extracts were concentrated to dryness under reduced pressures at 40 °C using a rotary evaporator and dissolved in methanol to make a stock solution of 50 mg/ml and used for different antioxidant activities. The effect of the extracts on DPPH (1,1-diphenyl-2-picrylhydrazyl) radical was determined following the standard method described by Liyana- Pathiranan and Shahidi (2005). The quantity of DPPH radical scavenged was calculated using the following equation:

DPPH radical scavenging activity (%) = [(AbsControl–AbsSample)]/(Abs Control)]x100....Eqⁿ... (1)

| Abs | Control | = | absorbance | of | DPPH |
|-------------------|---------|---|------------|----|------|
| radical+methanol; | | | | | (2) |

| Abs | Sample | = | absorbance | of | DPPH |
|--------|-----------|-------|--------------|----|------|
| radica | al+sample | extra | ct/standard. | | (3) |

Where Abs = Absorbency

2.5. Statistical Analysis

The results obtained were subjected to statistical analysis as mean and standard deviation (Zar, 1984). The mean values and standard deviations were calculated from the data obtained from three different experiments. The statistical difference at p < 0.05 was considered to be significant.

3.Results and discussions 3.1. Nutritional content

In the present study, a comparative analysis of nutritional composition between the raw and boiled tubers has been carried out, and the proximate content of the raw and boiled tubers is presented in Table-1. The moisture content of the raw tuber was found to be relatively low (74.89±0.54%) compared to the boiled tuber $(80.48\pm1.18\%)$. The ash content of the raw tuber was found to be high $(2.57\pm0.04\%)$ compared to the boiled tuber $(1.66\pm0.34\%)$. We found a very low amount of fats in both raw tubers $(0.19\pm0.01\%)$ and boiled tuber $(0.14\pm0.012\%)$, which are not significantly different. Total Carbohydrate (31.62±0.46%) and reducing sugar content $(0.018\pm0.008\%)$ of raw tuber was found to be relatively high compared to the boiled tuber (23.94±0.50% and 0.012±0.008%). In contrast, it was found that the boiled tuber contained an albeit high amount of starch

(11.67±0.65%) compared to the raw tuber $(8.6\pm0.54\%)$. The protein content of the raw and boiled tuber was found to be 3.48±0.92% and 2.25±0.16%, respectively. The total free amino acid content was found to be slightly high in the raw tuber $(1.45\pm0.05\%)$ compared to the boiled tuber (0.59±0.13%). HPLC analysis of both the raw and boiled tubers for different amino acids revealed that out of nine essential amino acids. six amino acids (Histidine, methionine, lysine, phenylalanine, threonine, valine) are observed in the raw tuber and four amino acids (lysine, threonine, valine, and phenylalanine) in the boiled tuber (Table 1). Phenylalanine was present in the highest amount in both raw and boiled tuber, followed by valine. Histidine, methionine, and cysteine amino acids were found in minimum quantity in the raw tuber and were not detected in the boiled samples. The observations of the mineral compositions of the raw and boiled tubers are presented in Table 1. The sodium, potassium, phosphorus, iron, and calcium content of the raw tubers were observed in the range of 316±27.78, 677.33±21.38, 153.20±17.17, 6.16±0.89, 290±4.13 mg/100g dry mass, respectively. In contrast, the estimated value of the above mineral contents in boiled were 119.36±16.25, 232.33±12.50, tuber 60.43±1.72, 3.24±1.06, 180±2.28 mg/100g dry mass, respectively. The other essential elements such as magnesium, zinc, manganese, and copper were found to be 203 ± 6.42 , 0.45 ± 0.95 , 4.2±2.16, and 0.79±0.62 mg/100g dry mass, respectively in the raw tuber. In contrast, the boiled tuber contained a lower amount of magnesium, zinc, manganese, and copper elements (102±4.14, 0.18±0.83, 0.89±1.86, and 0.12±0.37 mg/100g dry mass, respectively).

| Table 1.Nutritional composition and Nutrient content | Raw tuber | Boiled tuber |
|---|--------------------|--------------------|
| | Proximate | |
| Moisture (%) | 74.89±0.54 | $80.48{\pm}1.18$ |
| Ash (%) | 2.57±0.04 | 1.66±0.34 |
| Total carbohydrate (%) | 31.62±0.46 | 23.94±0.50 |
| Starch (%) | 8.6±0.54 | 11.67±0.65 |
| Reducing sugar (%) | 0.018±0.008 | $0.012{\pm}0.008$ |
| Fat (%) | 0.19±0.01 | 0.14±0.012 |
| Protein (%) | 3.48±0.92 | 2.25±0.16 |
| Free amino acid (%) | 1.45±0.05 | 0.59±0.13 |
| Amino acid quantity (mg/100g) | | |
| Glutamic acid | 14.27±0.51 | 10.24±0.35 |
| Glutamine | 8.95±0.11 | 7.6±0.29 |
| Histidine | 1.033±0.19 | Nd |
| Arginine | 3.9±0.23 | 3.34±0.15 |
| Alanine | 5.2±0.15 | 3.29±0.32 |
| Serine | 6.54±0.03 | 5.85±0.25 |
| Tyrosine | 4.18±0.17 | 3.58±0.24 |
| Cysteine | 0.37±0.13 | Nd |
| Methionine | 1.16±0.03 | Nd |
| Proline | 4.01±0.01 | $3.46{\pm}0.03$ |
| Glysine | 5.5±0.03 | 6.3±0.08 |
| Lysine | 3.54±0.43 | $2.32{\pm}0.07$ |
| Threonine | 4.42±0.27 | 2.05±0.05 |
| Valine | 5.19±0.11 | 4.43±0.03 |
| Phenylalanine | 5.29±0.35 | 4.83±0.21 |
| Mineral element (mg/100g) | | |
| Sodium | 316±27.8 | 120±16.3 |
| Potassium | 677±21.4 | 232±12.5 |
| Phosphorus | 153±17.2 | 60.4±1.72 |
| Iron | 6.16±0.89 | 3.24±1.06 |
| Calcium | 290±4.13 | 180±2.28 |
| Magnesium | 203±6.42 | 102±4.14 |
| Zinc | 0.45±0.95 | 0.18±0.83 |
| Manganese | 4.2±2.16 | 0.89±1.86 |
| Copper | 0.79±0.62 | 0.12±0.37 |
| Vitamin mg/100g | | |
| Vitamin C | 99.5±0.94 | 70.7±1.19 |
| Vitamin B1 | $0.007{\pm}0.0008$ | 0.005 ± 0.0004 |
| Vitamin B2 | $0.027{\pm}0.007$ | $0.014{\pm}0.003$ |
| Vitamin B3 | 27.38±1.42 | 14.65±1.25 |
| Vitamin B6 | 0.128±0.028 | $0.084{\pm}0.006$ |

Table 1. Nutritional composition analysis of raw and boiled tubers of *Dioscorea bulbifera*

Not detected

The details of vitamin analysis of Dioscorea bulbifera are shown in Table 1. The ascorbic acid content of the raw tuber was found to be 99.5±0.94 mg/100g dry mass. In contrast, the ascorbic content of the boiled tuber was found to be 70.7 ± 1.19 mg/100g dry mass. The content of vitamin B1, vitamin B2, vitamin B3, and vitamin B6 for the raw tuber was found to be 0.007±0.0004, 0.027±0.007, 27.38±1.42, and 0.128±0.028 mg/100g dry mass, respectively. In contrast, the boiled tuber's vitamin B1, vitamin B2, vitamin B3, and vitamin B6 content was found to be comparatively in lesser quantity than that of the raw tubers (Table 1). Among all the contents analyzed, it has been observed that in comparison to the present finding, a higher amount of fat, starch, protein, potassium, calcium, zinc, and copper has been reported earlier by other researchers (Polycarp et al., 2012). In contrast, carbohydrate and sodium contents reported earlier have been lower than the present findings (Sanful et al., 2013). A very similar amount of ash, protein, phosphorus, manganese, magnesium, ascorbic, and vitamin contents has been observed compared to the earlier report (Okwu and Ndu, 2006). In comparison to the boiled tuber, raw tubers are found rich in nutrition (Table-1) which might be due to the leaching and degradation of most of the vitamins and mineral contents during boiling. In comparison to the previous reports, the variation observed in the nutritional content of the tuber in the current report might be due to the soil Physico-chemical properties, soil fertility, geographical and climatic condition, genetic variation, etc. (Bhandari and Kawabata, 2004). Overall the results revealed that the tuber of Dioscorea bulbifera are good sources of minerals and could be used as food supplements

3.2. Bioactive compounds

Bioactive compounds are secondary metabolites of the plant that have pharmacological or toxicological effects in humans and animals. Tannin is one of the phenolic compounds which gives an astringent and bitter taste. Tannins act as antidiarrheal, haemostatic and anti hemorrhoidal, anti-

inflammatory. and antibacterial. antiviral antiseptic, antioxidant. Diosgenin content plants are grown for steroid preparation (Behera et al., 2010). Saponins are responsible for the reduction of cholesterol levels in animals along with other animals. (Desai et al., 2017). The analysis showed that raw tuber contained 160.2 ± 0.84 mg/100g while boil tuber contained 12.5±0.11mg/100g of diosgenin. Tannin content was found to be 180.11±0.32 mg/100g and 12.09±0.12mg/100g for raw and boil tuber, respectively, in this present study. The saponin content of raw and boil tuber was found to be 150.34±0.67mg/100g and 21.26mg/100g, respectively. Ghosh et al. (2014) reported diosgenin exhibited potent inhibition against both porcine pancreatic alpha-amylase and alpha-glucosidase as well as against crude murine amylase and glucosidase and acts as lead candidate in managing Type II Diabetes Behera et al. (2010) reported Mellitus. 1383mg/100g of diosgenin for Dioscorea bulbifera tuber. Polycarp et al. (2012) reported 10.98mg/100g of tannin in Dioscorea bulbifera. Princewill-Ogbonna and Ibeji (2015) found 8.49- 14.03mg/100g of saponin content of three cultivars of Dioscorea bulbifera.

3.3. Antioxidant properties

Phenols and flavonoids are compounds having antioxidant activity hence can absorb and neutralize free radicals, quenching singlet and triplet oxygen, or decompose peroxides (Luis et al., 2012). Among boiled and raw tuber, the latter one showed excellent radical scavenging activity, which is significantly higher than that of ascorbic acid and BHT (Butvlated Hydroxytoluene). Methanolic extracts of raw and boiled tubers of Dioscorea bulbifera were used in the present study to determine the DPPH radical scavenging activity, and results were compared with standard ascorbic acid and BHT (Figure 1). The IC_{50} value (the concentration of the sample that reduced 50% of the absorbances of DPPH) of raw and boiled tuber, ascorbic acid, and BHT are included in Figure 2. Higher the IC₅₀ value signifies less antioxidant activity and vice-versa. It was found that the methanolic

extract of the raw tuber has significantly higher antioxidant activity (IC₅₀ value is 46.11 μ g/ml) compared to the ascorbic acid (IC₅₀ value is 92.86 µg/ml), BHT (IC₅₀ value is 54.35 µg/ml) and boiled tuber extract (IC₅₀ value is 455.37 µg/ml). Phenolic content of the raw tuber extracted with different solvents such as acetone, methanol, and water was found to be 95.92±11.78, 121.11±13.71, and 205.19±15.91 mg GAE/100g dry weight, respectively (Table 2). In contrast, the boiled tuber's acetone, methanol, and water extract contained 12.71 \pm 5.28, 23.04 \pm 5.60, and 54.18 \pm 10.40 mg GAE/100g dry weight, respectively of total phenols. Previously, Bhandari et al. (2003) have reported 166 mg GAE/100g fresh weight of total phenols for the acetone extract of Dioscorea bulbifera tuber. Total flavonoid content with the methanol, acetone, and water extract of raw tuber was 359.82±18.10, 232.10±34.22, and 387.71±9.96 mg quercetin equivalent/100 g dry weight (Table 2). In contrast, the methanol,

acetone, and water extract of boiled tuber possessed a quite low flavonoid content viz. 75.91±12.63, 54.83±7.11, and 64.13±11.04 mg quercetin equivalent/100 g dry weight. Okwu and Ndu (2006) reported 8.04 mg quercetin equivalent /100g, and Adeosun et al. (2016) reported 5.36 mg quercetin equivalent/100g of flavonoid contents of Dioscorea bulbifera tuber on the dry weight basis. TPC (total phenol content) of tuber positively correlated with DPPH scavenging activity at the 0.01 level (R²=0.9762) (Figure 3). TFC (total flavonoid content) and DPPH also positively correlated at 0.01 level ($R^2=0.9717$) (Figure 4). The result of the analysis showed phenols are potent antioxidant and scavenging agents.

Overall, for treating radical-related pathological damage, the tuber of *Dioscorea bulbifera* can be used as a therapeutic agent. The current report also suggests that in *Dioscorea bulbifera*, polyphenol is essential component responsible for its antioxidant activities.

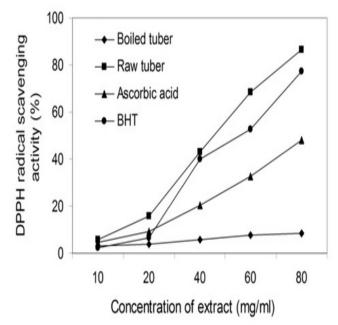


Figure 1. Comparison of DPPH radical scavenging activity of methanolic extract of the raw and boiled tuber of *Dioscorea bulbifera* with ascorbic acid and BHT.

| Polyphenol | olyphenol Raw tuber | | | | Boiled tuber | | | |
|------------|---------------------|-------------|-------------|------------------|------------------|-------------|--|--|
| content | Methanol | Acetone | Water | Methanol | Acetone | Water | | |
| | extract | extract | extract | extract | extract | extract | | |
| Phenols | 121.1±13.71 | 95.92±11.78 | 205.2±15.91 | 23.04 ± 5.60 | 12.71 ± 5.28 | 54.18±10.40 | | |
| (mg/100 g | | | | | | | | |
| dry mass) | | | | | | | | |
| Flavonoid | 359.8±18.10 | 232.1±34.22 | 387.7±9.96 | 75.91±12.63 | 54.83±7.11 | 64.13±11.04 | | |
| Mg/100 g | | | | | | | | |
| dry mass) | | | | | | | | |
| DPPH | | | | | | | | |

Table 2. Phenols and flavonoid content of raw and boiled tubers of Dioscorea bulbifera

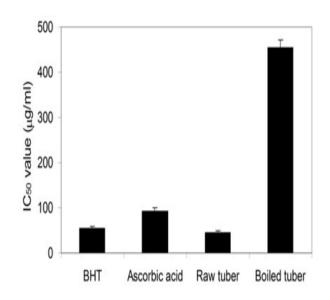


Figure 2. The IC₅₀ value of raw and boiled tuber, ascorbic acid, and BHT

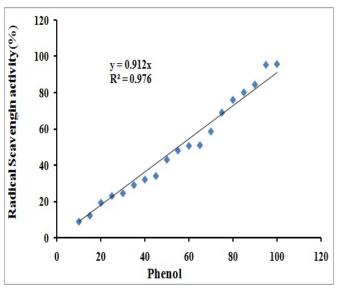


Figure 3. Correlation between TPC and DPPH

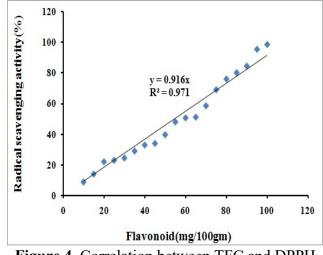


Figure 4. Correlation between TFC and DPPH

4. Conclusions

We have the nutritional evaluated composition and antioxidant activity of the tuber of D. bulbifera, both in raw and in boiled form. We found that the nutritional composition of the raw form is very rich than the boiled form of the tubers. The antioxidant activity of the raw tuber was found to be significantly very high compared to the ascorbic acid and BHT. The total phenolics and flavonoids are found to be significantly high in both raw and boiled tubers. present also The study emphasizes phytochemical analysis. Hence the tuber D. bulbifera could be a good candidate for functional foods.

5. References

- Abara, A.E. (2011). Proximate and mineral elements composition of the tissue and peel of *Dioscorea bulbifera* tuber. *Pakistan Journal of nutrition*, 10, 543-551.
- Adeosun, O.M., Arotupin, D.J., Toba, O.A., Adebayo, A.A. (2016). Antibacterial activities and phytochemical properties of extracts of *Dioscorea bulbifera* Linn (Air potatoe) tubers and peels against some pathogenic bacteria. *Journal of phytopharmacology*, 5, 20-26.
- Behera, K.K., Sahoo, S., Prusti, A. (2010). Biochemical quantification of diosgenin and ascorbic acid from the tubers of different Dioscorea species found in Orissa. *Libyan*

Agriculture Research Center Journal International, 2, 123-127.

- Bhandari, M.R., Kawabata, J. (2004). Organic acid, phenolic content and antioxidant activity of wild yam (Dioscorea spp.) tubers of Nepal. *Food Chemistry*, 88, 163-168.
- Bhandari, M.R., Kasai, T. and Kawabata, J. (2003). Nutritional evaluation of wild yam (Dioscorea spp.) tubers of Nepal. *Food Chemistry*, 82, 619-623.
- Ghosh, S., Derle, A., Ahire, M., More, P., Jagtap, S., Phadatareet, S.D., Patil, A.B., Jabgunde, A.M., Sharma, G.K., Shinde, V.S., Pardesi, K., Dhavale, D.D., Chopade, B.A. (2013). Phytochemical Analysis and Free Radical Scavenging activity of Medicinal Plants *Gnidia glauca* and *Dioscorea bulbifera*. *PloS One*, 8(12), 1–18.
- Ghosh, S., Jagtap, S., More, P., Shete, U.J., Maheshwari, N.O., Rao, S.J., Kitture R., Kale, S., Bellare, J., Patil S, Pal, J.K., Chopade B.A. (2015). *Dioscorea bulbifera* Mediated Synthesis of Novel AucoreAgshell Nanoparticles with Potent Antibiofilm and Antileishmanial Activity. *Journal of Nanomaterials*, 16(1), 1-12.
- Ghosh, S., More, P., Derle, A (2014). Diosgenin from Dioscorea bulbifera : Novel hit for treatment of type II Diabeteus mellitus with inhibitory activity against α -amylase and α -Glucosidase. *PloS One*, 9(9), 1-9

- Kalra, Y.P. (1998). Handbook of Reference Method for Plant Analysis, CRC Press, Taylor & Francis group, Boca Raton, FL.
- Kumar, S., Parida, A.K., Jena, P.K. (2013).
 Ethno-Medico-Biology of Ban-Aalu (Dioscorea species): A neglected tuber crops of Odisha, India. *International Journal of Pharmacy and Life Science*, 4, 3143-3150.
- Liyana-Pathiranan, C.M., Shahidi, F. (2005). Antioxidant activity of commercial soft and hard wheat (*Triticum aestivum* L) as affected by gastric pH conditions. *Journal of Agricultural and Food Chemistry*, 53, 2433-2440
- Lowery, O.H., Rosebrough, N.J., Farr, A.L., Randall, R.J. (1951). Protein measurement with the Folin phenol reagent. *Journal of Biological Chemistry*, 193, 265-275.
- Luis, A., Gil, N., Amaral, M.E., Duarte, A.P. (2012). Antioxidant activities of extracts from acacia melanoxylon, acacia dealbata and olea europaea and alkaloids estimation. *International Journal of Pharmacy and Pharmaceutical Science*, 4, 225-231.
- Murugan M., Mohan V.R. (2012). In vitro antioxidant studies of Dioscorea esculenta (Lour).Burkil. *Asian pacific Journal of tropical medicine*, 2, 1620-1624.
- Obdoni B.O., Ochuko, P.O. (2001). Phytochemical studies and comparative efficacy of the crude extracts of some Homostatic plants in Edo and Delta States of Nigeria. *Global Journal of Pure and Applied Science*, 8, 203-208.
- Okwu, D.E., Ndu, C.U. (2006). Evaluation of the phtonutrients, mineral and vitamin contents of some varieties of yam (Dioscorea spp.). *International Journal of Molecular Medicine and Advance Science*, 2, 199-203
- Ordon E.A., Gomez J.D., Vattuone M.A., Isla, M.I. (2006). Antioxidant activities of Sechium edule (Jacq.) Swart extracts. Food Chemistry, 97, 452-458.
- Osman, H. (1990). Dietary fiber composition of common vegetables and fruits in Malaysia. *Food Chemistry*, 37, 21-26.
- Padhan, B., Nayak, J.K., Panda, D. (2020). Natural antioxidant potential of selected

underutilized wild yams (Dioscorea spp.) for health benefit. *Journal of Food Science and Technology*, 57, 2370–2376.

- Pal, P., Kaur, P., Singh, N., Kaur, A., Misra, N.N., Tiwari, B.K. (2016). Effect of nonthermal plasma on physico-chemical, aminoacid composition, pasting and protein characteristics of short and long grain rice flour. *Food Research International*, 81, 50-57.
- Pallab, K., Tapan, B., Tapas, P., Ramen, K. (2013). Estimation of total flavonoids content (TPC) and antioxidant activities of methanolic whole plant extract of *Biophytum sensitivum Linn. Journal of Drug Delivery and Therapeutics*, 3(4): 33-37.
- Perales, S., Barbera, R., Lagarda, M.J., Farre, R. (2005). Bioavilability of calcium from milk based formulas and fruit juices containing milk and cereals estimated by in vitro methods(solubility, dialysability and uptake and transport by caco-2 cells). *Journal of Agricultural and Food Chemistry*, 53, 3721-3726.
- Polycarp, D., Afoakwa, E.O., Budu, A.S., Otoo, E. (2012). Characterization of chemical composition and anti-nutritional factors in seven species within the Ghanaian yam germplasm. *International Food Research Journal*, 19, 985-992.
- Ranganna, S.(2007). Handbook of analysis and quality control for fruits and vegetable products, 2nd edn. Tata McGraw-Hill Publishing Company Ltd, New York
- Sadasivam, S., Manickam, A. (2008). Biochemical methods, 3rd edn. New age international (P) Limited, Publishers, New Delhi
- Sanful, R.E., Oduro, I., Ellis, W.O. (2013). Proximate and Functional properties of five local varieties of aerial yam (Dioscorea bulbifera) in Ghana. *Middle-East Journal of Scientific Research*, 14, 947-951.
- Schanderl, S.H. (1970). In:Method in Food Analysis, Academic Press, New York, p.709
- Thayumanavan, B., Sadasivam, S. (1984). Physicochemical basis for the preferential

uses of certain rice varieties. *Plant Foods for Human Nutrition*, 34, 253-259.

- Uematsu, Y, Hirata, K., Saito, K. (2000). Spectrophotometric Determination of Saponin in Yucca Extract Used as Food Additive. *Journal of AOAC International*, 83, 1451-1454.
- Zar, J.H. (1984). Biostatistical Analysis. Englewood Cliffs NJ, Prentic Hall. 5, 437-467.

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